

Ligand Conjugation of Chemically Exfoliated MoS₂

Stanley S. Chou,⁺ Mrinmoy De,⁺ Jaemyung Kim,⁺ Segi Byun,^{+,||} Conner Dykstra,⁺ Jin Yu,^{||} Jiaying Huang,^{*,+} and Vinayak P. Dravid^{*,+}

⁺Department of Material Science & Engineering, International Institute for Nanotechnology, Northwestern University, Evanston, Illinois 60208, United States

^{||}Department of Material Science and Engineering, Korea Advanced Institute Science and Technology, Yuseong-gu, Daejeon, Republic of Korea 305-701

Supporting Information

ABSTRACT: MoS₂ is a two-dimensional material that is gaining prominence due to its unique electronic and chemical properties. Here, we demonstrate ligand conjugation of chemically exfoliated MoS₂ using thiol chemistry. With this method, we modulate the ζ -potential and colloidal stability of MoS₂ sheets through ligand designs, thus enabling its usage as a selective artificial protein receptor for β -galactosidase. The facile thiol functionalization route opens the door for surface modifications of solution processable MoS₂ sheets.

The recent excitement over two-dimensional atomically layered materials has been fueled in large part by desires to exploit their low-dimensional properties, which can be distinct from their bulk counterparts. One of these emerging materials, MoS₂, has received a fair share of attention due to its applicability in areas ranging from catalysis¹ and electronics^{2–5} to biomedicine.⁶ Nevertheless, to fully harness the capabilities of such new materials, ligand conjugations were often necessary. For example, it is typically a crucial step for solution processing, which enables the usage of nanoparticles as functional assembly blocks in bioimaging,⁷ electronics,⁸ sensing,⁹ and photovoltaics.¹⁰ Previously, attempts to functionalize MoS₂ have been limited to bulk-like materials characterized by hydrophobic cleavage planes.^{11,12} Specifically, Tremel and co-workers¹³ have modified hydrophobic MoS₂ particles using nickel-nitrioltriacetic acid chelation.¹³ However, colloidal surface modification of water dispersible, chemically exfoliated MoS₂ (ce-MoS₂) sheets has not been demonstrated.

To obtain large quantities of single-layer MoS₂ sheets, solution based exfoliation methods are often used. Most typically, this involves chemical exfoliation using lithium intercalation, which can produce single layer MoS₂ sheets in scalable quantities.^{14–16} In this method, lithium is inserted between MoS₂ layers and then reacted with water to produce hydrogen gas at the interface. This, in combination with ultrasonication, then enables the high yield production of water dispersible ce-MoS₂ sheets.¹⁴ Because of the violent nature of this reaction, however, MoS₂'s crystal structure becomes deformed,^{3,17} and internal edges (tears, pinholes and defects) can become visible.¹⁸ Previous work has suggested these edge sites to possess higher molecular affinities,¹⁹ with theoretical suggestions of thiol edge absorption.²⁰ It is thus possible that

ce-MoS₂, which possesses defects in both internal edges and perimeter edges, may be amenable to thiol ligand modifications. As this has not been explicitly demonstrated, we show conjugation of ce-MoS₂ sheets by thiol-terminated ligands (Figure 1, Supporting Information S1–S5). This was used to

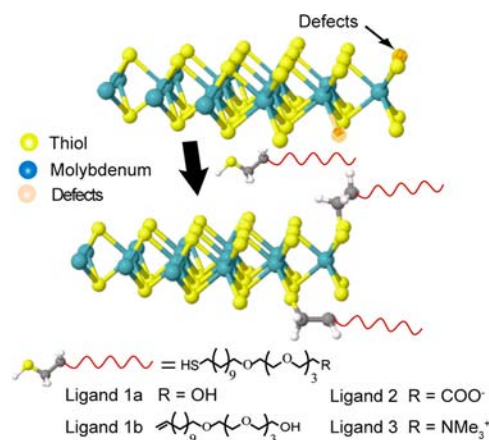


Figure 1. Structural models illustrating ligand conjugation of ce-MoS₂ sheets.

conveniently tune ce-MoS₂ and its biomolecular interactions. This route opens pathways for making solution processable MoS₂ with tunable colloidal properties and can potentially facilitate improvements in its processing.

In this report, the as-made ce-MoS₂ was first purified using exhaustive dialysis (7 days under continuous water flow). Survey of the material using X-ray Diffraction (XRD) showed primary peak at $2\theta = 14$, thus indicating absence of mixed lithium phase (Figure 2a).^{3,21} This was confirmed using Inductive Coupled Plasma-Mass Spectrometry (ICP-MS), which showed molecular ratios of samples obtained through dialysis to contain more than 10-fold less Li than the typical centrifugation purification process (vide infra). Atomic Force Microscopy (AFM) revealed sheets with average diameters of 0.7 μ m, and average thicknesses between \sim 0.8 and 1.5 nm, consistent with reported values of monolayers.²² Additionally, the internal edges mentioned previously become visible (Figures 2b, S6).¹⁸ Transmission Electron Microscopy (TEM)

Received: November 5, 2012

Published: March 11, 2013

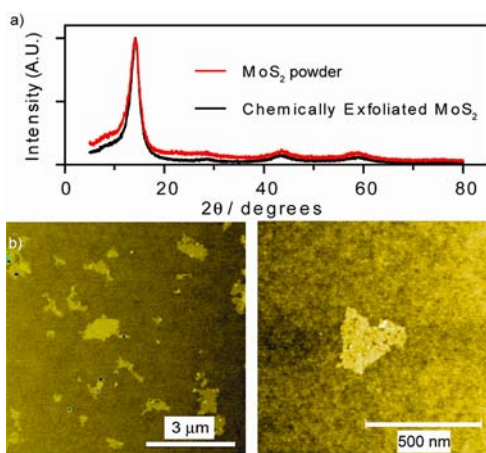


Figure 2. (a) XRD spectra of ce-MoS₂. (b) Low and high magnification AFM micrograph of ce-MoS₂.

Selective Area Diffraction (SAD) showed 6-fold symmetrical diffraction spots (2H phase) with additional points in between those diffraction points (1T phase, Figure S7). This is consistent with previous reports by Dungey et al.¹⁷ for ce-MoS₂, thus indicating a mixed 1T and 2H material. Separately, Dynamic Light Scattering (DLS) show native ce-MoS₂ sheets to have a ζ -potential of -50 mV (Figure 3a). To this end, there

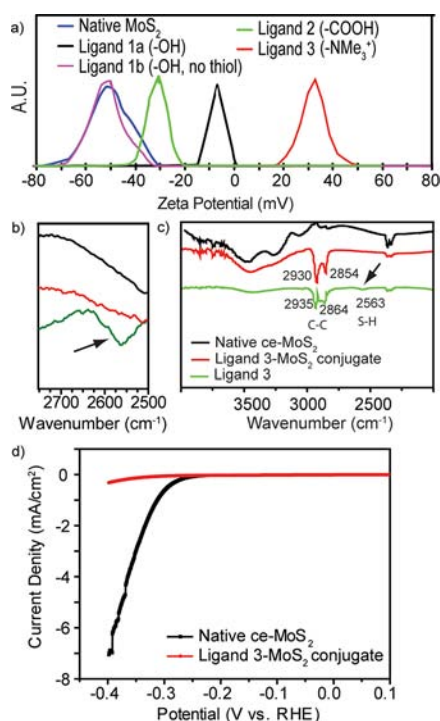


Figure 3. (a) ζ -potential, (b) FT-IR spectra focused upon the thiol peak, (c) FT-IR spectra showing all peaks, and (d) HER polarization curve before and after ligand conjugation.

are different models in literature regarding the origin of this negative charge. Principally, Divigalpitiya et al.¹⁹ proposed a model with charges at edges. Separately, Heising et al.²³ have a model based on partially oxidized Mo atoms on the basal plane due to Li intercalation. However, both agree that the sheets are defective, thus there is the possibility of chemical modification.

To illustrate this process, we first sought to understand the ligand affinity of chemically exfoliated MoS₂. For this, we synthesized a series of polyethylene glycol (PEG) ligands with head-groups of different charge (Figures 1, S1–S5). By including and excising the thiol moiety, we show that the thiol group is responsible for the ce-MoS₂ modification (**ligand 1a** vs **1b**).

We first examined the conjugation results using **ligand 1a** and **ligand 1b**. As can be seen in Figure 1, these ligands are nearly identical except for the thiol group on **ligand 1a**. Despite their similarities, only the ce-MoS₂ samples incubated with **ligand 1a** showed significant ζ -potential shifts after purification. The resultant ζ -potential of -7 mV represents a 43 mV shift from native ce-MoS₂, and is consistent with a neutral PEG functionalized colloid.¹⁹ Comparatively, **ligand 1b** produced no ζ -potential change, indicating no modification (Figure 3a). Because of the attenuated ζ -potential, **ligand 1a** conjugate was found to be less stable in water.

To expand upon the conjugations, **ligand 2** and **ligand 3** were synthesized. Here, the conjugates also show appropriate changes in ζ -potential as well, with **ligand 2** producing a ζ -potential of -29 mV and **ligand 3** a ζ -potential of $+36$ mV. Overall, **ligand 3** conjugates exhibited the greatest colloidal stability. We believe this to be attributed to the pH independent nature of the charged NMe₃⁺ group on **ligand 3**.

Further characterizations were then performed. First, Fourier Transform Infrared spectroscopy (FT-IR, Figures 3b,c, S9) revealed the exposed S–H band from free ligands at 2563 cm⁻¹. This peak becomes absent after conjugation with ce-MoS₂, giving indication that the thiol moiety becomes buried on to the ce-MoS₂ surface (Figure 3b).^{24,25} Presence of the ligands on the conjugates was also validated by monitoring the C–H aliphatic bands at 2854 and 2930 cm⁻¹. Here, these aliphatic bands appear on the conjugates at 2864 and 2935 cm⁻¹ (Figure 3c).²⁶ Unique pegylated ligand signature on conjugated ce-MoS₂ surfaces were also verified using X-ray Photoelectron Spectroscopy (XPS). For example, ether (C–O–C) bonds characteristic of PEG ligands appeared in all conjugated samples (286.6 eV, Figure S8). These results further corroborate presence of PEG ligands on the ce-MoS₂, and are consistent with thiol-PEG functionalized materials.^{27–29}

With the conjugation assessed, we then evaluate the effect of residual lithium (Li) from the exfoliation process. Here, ce-MoS₂ purified using the 3X centrifugation process described by Heising²³ produced a Li:Mo ratio of 0.25, consistent with their report. ce-MoS₂ purified using the dialysis process described here reduced the Li:Mo ratio to 0.02, more than 10-fold lower (Figure S10). However, the residual Li does not appear to effect conjugation results radically (Figure S11)

We also examined the effect of ligand conjugation on the hydrogen evolution reaction (HER) process. As previous studies suggest edges to dominate HER catalysis,³⁰ we expect that ligand conjugation to suppress HER activity. Indeed, decrease catalytic activity was clearly observed (Figures 3c, S12). It thus appears that the thiol ligands can at minimum functionalize the various edges in ce-MoS₂.

We then monitored the changes in colloidal stability with ligand conjugation. Noticeably, native ce-MoS₂ irreversibly precipitated due to restacking within the first 21 days (Figure 5a).¹⁴ Comparably, the conjugates maintained colloidal stability (**ligand 3**), or can be readily redispersed with shaking (**ligand 1a**, **2**) (Figure 4a). This is likely due PEG ligand preventing complete restacking. Because of the pH independent nature of

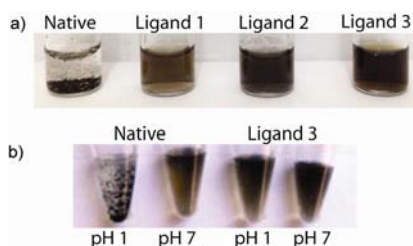


Figure 4. (a) Colloidal stability of ce-MoS₂ samples after 21 days in water. (b) Colloidal stability of ce-MoS₂ functionalized with NMe₃ terminated thiol at various pH.

the NMe₃⁺ group on **ligand 3**, its conjugates are also capable of resisting flocculation in acidic media (Figure 4b). To compare, native ce-MoS₂ flocculated at low pH.¹⁴ **Ligand 3** conjugates, on the other hand, were able to maintain colloidal stability (Figure 4b).

The combination of ζ -potential tailoring and pegylation also enables the tuning of ce-MoS₂ for selective host–guest interactions with biomolecules. As we have recently shown ce-MoS₂ to be a promising photothermal agent,³¹ ligand modifications shown here can have immediate applications. To demonstrate this, we show the ligands ability to change ce-MoS₂'s function as an artificial protein receptor. To this end, we use β -galactosidase (β -gal), a hydrolase enzyme. Here, β -gal, due to a ring of anionic residues (Asp and Glu) surrounding its active site, does not interact with native ce-MoS₂ (Figure Sa).^{32,33} By tuning ce-MoS₂ for the correct electrostatic host–

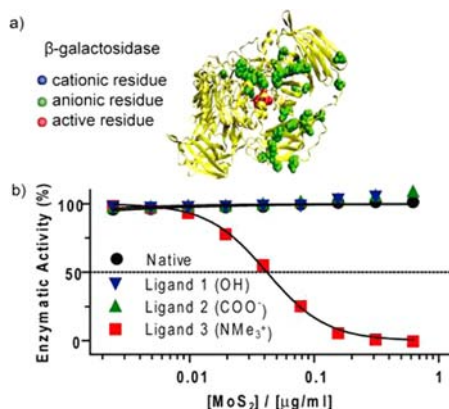


Figure 5. (a) Structure of a single unit of a β -Gal tetramer. Anionic residues (Asp and Glu) surrounding β -gal's active site are highlighted (bottom) (b) Concentration dependence of β -gal inhibition with various ce-MoS₂ conjugate.

guest interactions, we thus enable selective complexation and inhibition of this enzyme.³⁴ For this, 62.5 pM β -gal was incubated with various concentrations of ce-MoS₂-conjugates. After 30 min, *o*-nitrophenyl- β -D-galactopyranoside (ONPG), a chromogenic substrate, was added to monitor β -gal activity. From this experiment, it can be seen that only the **ligand 3** conjugate, which is cationic, showed ability to modulate β -gal activity. Specifically, 50% of β -gal activity can be inhibited by 0.04 μ g/mL of the conjugate (Figure 5b). These results demonstrate the possibility to tune ce-MoS₂ for selective host–guest interactions with enzymes. Further development of this demonstrated concept can thus be utilized for biosensor design.^{9,35,36}

In summary, we have demonstrated ligand conjugation of chemically exfoliated MoS₂ sheets through thiol chemistry. In this manner, we were able to tune the ζ -potential and surface functionality of ce-MoS₂ sheets to enable its broad usage as artificial receptors for enzymes. Because these aforementioned assemblies have potential applications in sensing,^{35,37} energy,^{1,30} and so forth, and because devices using 2D materials can outperform their nanoparticles,^{34,38} it is possible that the functionalized ce-MoS₂ assemblies may enjoy similar advantages. The ability to functionalize these sheets through facile thiol chemistry suggests that similar chemistry may be applied to other chemically exfoliated transition metal dichalcogenide sheets (Figure S11), and can therefore open the window for further applications and solution processing of these emerging two-dimensional materials.

■ ASSOCIATED CONTENT

📄 Supporting Information

Synthetic protocols, experimental procedures, and additional characterizations. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

v-dravid@northwestern.edu; jiaxing-huang@northwestern.edu

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was partly funded by the National Cancer Institute Center for Cancer Nanotechnology Excellence (CCNE) initiative at Northwestern University Award No. U54CA119341. J.H. acknowledges support from National Science Foundation (DMR CAREER 0955612) and the Alfred P. Sloan Research Foundation. S.B. was supported by Basic Science Research Program through the National Research Foundation of Korea funded by the MEST (Project no. 2012-000181). We thank NU-HTA and NU NUANCE and NU-QBIC for characterization equipment availability and expertise. We also thank A. Yan and Y. Huang for assistance with EM microscopy and M. Rotz and Dr. K. MacRenaris for assistance with ICP-MS. S.S.C. thanks US DHS for a graduate fellowship.

■ REFERENCES

- (1) Li, Y.; Wang, H.; Xie, L.; Liang, Y.; Hong, G.; Dai, H. *J. Am. Chem. Soc.* **2011**, *133*, 7296.
- (2) Radisavljevic, B.; Radenovic, A.; Brivio, J.; Giacometti, V.; Kis, A. *Nat. Nanotechnol.* **2011**, *6*, 147.
- (3) Eda, G.; Yamaguchi, H.; Voiry, D.; Fujita, T.; Chen, M.; Chhowalla, M. *Nano Lett.* **2011**, *11*, 5111.
- (4) Zhang, H.; Loh, K. P.; Sow, C. H.; Gu, H.; Su, X.; Huang, C.; Chen, Z. K. *Langmuir* **2004**, *20*, 6914.
- (5) Liu, K.-K.; Zhang, W.; Lee, Y.-H.; Lin, Y.-C.; Chang, M.-T.; Su, C.-Y.; Chang, C.-S.; Li, H.; Shi, Y.; Zhang, H.; Lai, C.-S.; Li, L.-J. *Nano Lett.* **2012**, *12*, 1538.
- (6) Wu, H.; Yang, R.; Song, B.; Han, Q.; Li, J.; Zhang, Y.; Fang, Y.; Tenne, R.; Wang, C. *ACS Nano* **2011**, *5*, 1276.
- (7) Medintz, I. L.; Uyeda, H. T.; Goldman, E. R.; Mattoussi, H. *Nat. Mater.* **2005**, *4*, 435.
- (8) Nakanishi, H.; Walker, D. A.; Bishop, K. J. M.; Wesson, P. J.; Yan, Y.; Soh, S.; Swaminathan, S.; Grzybowski, B. A. *Nat. Nanotechnol.* **2011**, *6*, 740.
- (9) De, M.; Rana, S.; Akpınar, H.; Miranda, O. R.; Arvizo, R. R.; Bunz, U. H. F.; Rotello, V. M. *Nat. Chem.* **2009**, *1*, 461.

- (10) Yaacobi-Gross, N.; Soreni-Harari, M.; Zimin, M.; Kababya, S.; Schmidt, A.; Tessler, N. *Nat. Mater.* **2011**, *10*, 974.
- (11) Rapoport, L.; Fleischer, N.; Tenne, R. *J. Mater. Chem.* **2005**, *15*, 1782.
- (12) Yarar, B.; Kaoma, J. *Colloids Surf.* **1984**, *11*, 429.
- (13) Tahir, M. N.; Zink, N.; Eberhardt, M.; Therese, H. A.; Kolb, U.; Theato, P.; Tremel, W. *Angew. Chem., Int. Ed.* **2006**, *45*, 4809.
- (14) Joensen, P.; Frindt, R. F.; Morrison, S. R. *Mater. Res. Bull.* **1986**, *21*, 457.
- (15) Schumacher, A.; Scandella, L.; Kruse, N.; Prins, R. *Surf. Sci.* **1993**, *289*, L595.
- (16) Zeng, Z.; Yin, Z.; Huang, X.; Li, H.; He, Q.; Lu, G.; Boey, F.; Zhang, H. *Angew. Chem., Int. Ed.* **2011**, *50*, 11093.
- (17) Dungey, K. E.; Curtis, M. D.; Penner-Hahn, J. E. *Chem. Mater.* **1998**, *10*, 2152.
- (18) Eda, G.; Fujita, T.; Yamaguchi, H.; Voiry, D.; Chen, M.; Chhowalla, M. *ACS Nano* **2012**, *6*, 7311.
- (19) Divigalpitiya, W. M. R.; Frindt, R. F.; Morrison, S. R. *Science* **1989**, *246*, 369.
- (20) Raybaud, P.; Hafner, J.; Kresse, G.; Toulhoat, H. *Phys. Rev. Lett.* **1998**, *80*, 1481.
- (21) Joensen, P.; Crozier, E. D.; Alberding, N.; Frindt, R. F. *J. Phys. C* **1987**, *20*, 4043.
- (22) Late, D. J.; Liu, B.; Matte, H. S. S. R.; Rao, C. N. R.; Dravid, V. P. *Adv. Funct. Mater.* **2012**, *22*, 1894.
- (23) Heising, J.; Kanatzidis, M. G. *J. Am. Chem. Soc.* **1999**, *121*, 11720.
- (24) Shi, W.; Sahoo, Y.; Swihart, M. T. *Colloid Surf., A* **2004**, *246*, 109.
- (25) Thangadurai, P.; Balaji, S.; Manoharan, P. T. *Nanotechnology* **2008**, *19*, 435708.
- (26) Porter, M. D.; Bright, T. B.; Allara, D. L.; Chidsey, C. E. D. *J. Am. Chem. Soc.* **1987**, *109*, 3559.
- (27) Harder, P.; Grunze, M.; Dahint, R.; Whitesides, G. M.; Laibinis, P. E. *J. Phys. Chem. B* **1998**, *102*, 426.
- (28) Wagner, C. D.; Muilenberg, G. E. *Handbook of X-ray Photoelectron Spectroscopy: A Reference Book of Standard Data for Use in X-ray Photoelectron Spectroscopy*; Perkin-Elmer Corp.: Physical Electronics Division, 1979.
- (29) Weisbecker, C. S.; Merritt, M. V.; Whitesides, G. M. *Langmuir* **1996**, *12*, 3763.
- (30) Jaramillo, T. F.; Jørgensen, K. P.; Bonde, J.; Nielsen, J. H.; Horch, S.; Chorkendorff, I. *Science* **2007**, *317*, 100.
- (31) Chou, S. S.; Kaehr, B.; Kim, J.; Foley, B. M.; De, M.; Hopkins, P. E.; Huang, J.; Brinker, C. J.; Dravid, V. P. *Angew. Chem., Int. Ed.* **2013**, DOI: 10.1002/anie.201209229.
- (32) Verma, A.; Simard, J. M.; Worrall, J. W. E.; Rotello, V. M. *J. Am. Chem. Soc.* **2004**, *126*, 13987.
- (33) Jacobson, R. H.; Zhang, X. J.; DuBose, R. F.; Matthews, B. W. *Nature* **1994**, *369*, 761.
- (34) De, M.; Chou, S. S.; Dravid, V. P. *J. Am. Chem. Soc.* **2011**, *133*, 17524.
- (35) Miranda, O. R.; Chen, H.-T.; You, C.-C.; Mortenson, D. E.; Yang, X.-C.; Bunz, U. H. F.; Rotello, V. M. *J. Am. Chem. Soc.* **2010**, *132*, 5285.
- (36) Miranda, O. R.; You, C.-C.; Phillips, R.; Kim, I.-B.; Ghosh, P. S.; Bunz, U. H. F.; Rotello, V. M. *J. Am. Chem. Soc.* **2007**, *129*, 9856.
- (37) Lee, J.-S.; Ulmann, P. A.; Han, M. S.; Mirkin, C. A. *Nano Lett.* **2008**, *8*, 529.
- (38) Chou, S. S.; De, M.; Luo, J.; Rotello, V. M.; Huang, J.; Dravid, V. P. *J. Am. Chem. Soc.* **2012**, *134*, 16725.